# A Basis of the Crenation of Erythrocyte Ghosts by Electrolytes

Pierre Wong\*

Laboratoire de Chimie des Protéines, Section Théorique, Montréal, Québec, Canada H2X 2B4

**Abstract:** Erythrocyte ghosts isolated in hemolytic hypotonic buffers, pH 7.0-8.0,  $0-4^{\circ}$ C were previously observed to be crenated by electrolytes. This shape transformation, which occurred rapidly and reversibly, was attributed to electrostatic interactions of cations with flexible filamentous anionic spectrin, the major protein component of the skeleton and/or anionic phospholipids, since divalent cationic salts crenated at concentrations substantially lower than those of monovalent cationic salts and that crenation appeared unrelated to the anion species. However, crenation by electrolytes was markedly influenced by ionic conditions and temperature. A mechanism of the erythrocyte shape control has been previously suggested in which band 3 (AE1), exchanging the monovalent anions Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> and linked to spectrin, plays a pivotal role. Briefly, the alternative recruitment of its inward-facing (band 3<sub>i</sub>) and outward-facing (band 3<sub>o</sub>) conformations contract and relax the skeleton, thereby promoting echinocytosis and stomatocytosis, respectively. Band 3 transports also other anions, including endogenous inorganic phosphate, but at a slow rate. This mechanism would explain the above observations and would lead to some inferences, one of which is a Cl<sup>-</sup>dependent crenation by Mg<sup>2+</sup> and Ca<sup>2+</sup>, suggesting that they specifically bind on sites on spectrin.

Keywords: Band 3, Buffer, Crenation, Electrolytes, Spectrin.

### **INTRODUCTION**

Ghosts or post-hemolytic plasma membranes of erythrocytes isolated free of hemoglobin in a hemolytic hypotonic buffer, pH 7.0-8.0, 0-4°C since the early 1960's may have a shape similar to or reminiscent of the resting biconcave disc shape and other familiar shapes of the erythrocyte, but seemingly influenced by the type of the buffer and the pH [1-9]. They are relatively well characterized with respect to their components and assemblies of these components [5] and can be made impermeable to small solutes by resealing under certain conditions in the presence of  $Mg^{2+}$  [1, 10, 11]. They have been used for multiple purposes, including monitoring transport of solutes across the membrane [1, 10, 11], probing membrane glucose metabolism and membrane fluctuations in the oxygenation and deoxygenation cycle, altering glucose metabolism and Mg<sup>2+</sup> level [12-14], testing hypotheses on the erythrocyte shape [2-8], [15-22] and as drug carriers and electrosensors [23-25]. Ghosts isolated in hemolytic hypotonic buffers, pH 7.0-8.0, 0-4°C were previously observed to be crenated by electrolytes (e.g. NaCl, NaCH<sub>3</sub>COO, CaCl<sub>2</sub>, SrCl<sub>2</sub>, MgSO<sub>4</sub>) [2-6] (see Table 1). This shape transformation was rapid and reversible [4, 6]. It was generally attributed to electrostatic interactions of cations with flexible filamentous anionic spectrin, the major protein component of the two-dimensional reticulated skeleton, apposed at the inner leaflet of the membrane lipid bilayer, and/or anionic phospholipids, mainly localized in the inner composed of phosphatidylserine leaflet and and phosphatidylinositides [3-5, 17], since divalent cationic salts crenated at concentrations substantially lower than those of monovalent cationic salts (0.1-0.2 mM versus 30 mM) [2-6], and that crenation appeared unrelated to the anionic species [3, 5]. However, this crenation was markedly influenced by ionic conditions and temperature, as indicated by the following observations: a) Ghosts in a 10 mM zwitterionic N-2hydroxyethylpiperazine-N'-2-ethanesulfonic acid (Hepes) buffer, pH 7.0, 4°C significantly crenated only in the presence of both 0.2 mM MgCl<sub>2</sub> and 10 mM NaCl [6]. b) Ghosts in 5 mM sodium phosphate (NaPi) buffer, pH 8.0, 0°C were strongly crenated by 1 mM MgSO<sub>4</sub>, but those in 25-30 mM MgSO<sub>4</sub> were uncrenated [5]. c) Ghosts in 8 different 10 mM anionic buffers, pH 7.4, 0°C containing 0.25 mM dithioerythritol were crenated by isotonic saline, and remained crenated after raising the temperature to 30°C, but were transformed into smooth discs and cups after the addition of adenosine triphosphate (ATP)-Mg<sup>2+</sup> complex [22]. On the other hand, ghosts in 14 different 10 mM zwittterionic buffers and 2 different 10 mM cationic buffers, pH 7.4, 0°C containing 0.25 mM dithioerythritol, having a lower ionic strength, were also crenated by isotonic saline, but were spontaneously transformed into smooth discs and cups when the temperature was elevated at 30°C. This spontaneous transformation was not reversed after washing these ghosts with anionic buffers, but ghosts isolated in anionic buffers underwent this spontaneous transformation after being washed with zwitterionic buffers; dithiothreitol appeared to be added in order to exclude the possibility that this spontaneous transformation was caused by oxidation of membrane proteins [6]. Lastly, the spontaneous transformation into discs and cups was minimal when ghosts were in zwitterionic or cationic buffers at pH in the range of 6.5-7.0 or above the pK<sub>a</sub> of the zwitterionic buffers at  $0^{\circ}C$  [22].

<sup>\*</sup>Address correspondence to this author at the LCP, Section Théorique, 3415 Aylmer, Montréal, Québec, Canada H2X 2B4; Tel: 514-849-1550; Fax: 514-982-0341; E-mail: pwong\_LCP@yahoo.ca

## Table 1. Observations of Electrolytes and Buffers on Ghost and Erythrocyte Shapes

Buffer	Electrolytes	[Electrolyte] (mM)	Crenation	Ref.
Hemolytic hypotonic buffer	(ghosts)			
5.6 mM Tris-HCl-40 mM Te	s-Tea, pH 7.4 <sup>ª</sup>			
	-	-	Monoconcave discs	[2]
	NH <sub>4</sub> Cl	50 <sup>b</sup>	+	
	MgCl <sub>2</sub>	1 <sup>b</sup>	+	
	NiCl <sub>2</sub>	0.4 <sup>b</sup>	+	
5 mM Tris-HCl, 7 mM NaCl	, pH 7.4			
	-	-	Discs, Cups	[3,4]
	CaCl <sub>2</sub>	1	+	
	MgCl <sub>2</sub>	1	+	
	SrCl <sub>2</sub>	1	+	
	NaCl	30	+	
5 mM NaP <sub>i</sub> , pH 8.0	L			I
	-	-	Plump bowls with one shallow dimple	[5]
	NaCl	150	+	
	NaCl, glycerol	150, 300	+	
	NH4HCO3	150	+	
	CaCl <sub>2</sub>	1	+	
25-30 mM MgSO4	MgSO <sub>4</sub>	1		
10 mM Hepes, pH 7.0	MgSO <sub>4</sub>	25-30	Uncrenated	[5]
	-	-	Spheres	[6]
	CaCl <sub>2</sub>	0.1-1	+	
	MgCl <sub>2</sub>	0.2	Some crenation	
	NaCl	10	Some crenation	
	MgCl <sub>2</sub> , NaCl	0.2, 10	+	
	NaCl or KCl	70	+	
Hemolytic isotonic medium	(ghosts)			
150 mM NH <sub>4</sub> HCO <sub>3</sub>	NH4HCO3	150	Globoid	[5]
300 mM Glycerol	-	-	Globoid	[5]
	NaCl	150	Globoid	
Buffered isotonic sucrose (er	ythrocytes)	I	I	
300 mM Sucrose, NaCl, Hep	es, pH 6.0			
^	NaCl, Hepes	0.52, 0.0175	Crenation ->	[34]
			Discocytosis ->	
			Cupping °	

a, Ghosts in 28 mM Tris-HCl buffer, pH 7.4 were diluted with 4 volumes of 50 mM Tes-Tea (N-tris(hydroxymethyl)-2-aminoethanesulfonic acid- triethylamine) buffer, pH 7.4; b, electrolyte concentration causing 50 % crenation; c, the rapid morphological transformation crenation -> discocytosis -> cupping occurred in different zwitterionic buffers.



**Fig. (1). Scheme of a mechanism of control of the erythrocyte shape.** The transmembrane glycoprotein band 3 (AE1), which exchanges Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> and is linked to spectrin, the major skeleton protein, by the intermediary of ankyrin R, plays a central role in the mechanism. The alternative recruitment of band 3<sub>i</sub> and band 3<sub>o</sub> by the exchange of Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> folds and unfolds spectrin, thus promoting echinocytosis and stomatocytosis, respectively. The band 3<sub>o</sub>/band 3<sub>i</sub> ratio is determined by the Donnan ratio ( $\mathbf{r} = Cl_i/Cl_o = HCO_{3-i}/HCO_{3-o} = H^+_o/H^+_i$ ), but this ratio is 0. 1 and 10 in the self-exchanges of Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> indicating that the latter are intrinsically echinocytogenic and stomatocytogenic, respectively.

A mechanism of control of the erythrocyte shape has been previously proposed in which the anion exchanger band 3 (AE1) plays a central role (see Fig. 1) [26]. Briefly, band 3 is a 90-100-kDa transmembrane glycoprotein which rapidly exchanges the monovalent anions Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> and is linked via ankyrin R to flexible anionic spectrin, the major protein component of the two-dimensional reticulated skeleton. It transports also endogenous P<sub>i</sub> and a wide variety of other inorganic and organic anions, including bulky ones, but at a slow rate [27]. The alternative recruitment of its inward-facing (band  $3_i$ ) and outward-facing (band  $3_o$ ) conformations by exchange of Cl and HCO<sub>3</sub> folds and unfolds spectrin (or contracts and relaxes the skeleton), thus promoting echinocytosis and stomatocytosis, respectively. The band  $3_0$ /band  $3_i$  equilibrium ratio increases with the increase of the Donnan equilibrium ratio of anions Cl<sup>-</sup> and  $HCO_3^-$  and  $H^+$  (r =  $Cl_i^-/Cl_o^-$  =  $HCO_3^-/HCO_3^-$  =  $H^+_o/H^+_i$ ), which is determined by hemoglobin and the major organic phosphate intermediate 2,3-bisphophoglycerate. Substrates slowly transported by band 3, preferentially inwardly and outwardly are echinocytogenic and stomatocytogenic, respectively. Recently, a process of echinocytosis by glucose depletion has been suggested based on this mechanism [28]. As was indicated, this process would be compatible with the reversal of ghost crenation by buffered isotonic saline by ATP-Mg<sup>2+</sup> complex, since ATP is hydrolyzed by a vanadatesensitive Mg<sup>2+</sup>-ATPase which would promote the stomatocytogenic outward transport of divalent P<sub>i</sub> with a hydrogen ion ( $H^+$ ) (or monovalent  $P_i$ ) by band 3 [18, 20-22]. The plausibility of this process would also be further supported by two previous observations. a) Ghosts isolated in 5 mM NaP<sub>i</sub> buffer, pH 8.0, 0°C were morphologically indistinguishable before and after ATP depletion of erythrocytes by glucose depletion in a buffered isotonic saline at 37°C, and appeared as plump bowls with a single shallow dimple [5]. b) Rabbit erythrocytes suspended in phosphate-buffered isotonic saline, pH 7.7 were crenated after a blood storage of 1 day, but had a biconvave disc shape after suspending them in phosphate-buffered isotonic saline, pH 8.5, a pH markedly opposing the transport of divalent  $P_i$  with a H<sup>+</sup> by band 3, since its pK<sub>a</sub> is 7.31-7.18, 0-40°C [29, 30]. We have examined whether the mechanism could be the basis of the marked influence of the ionic composition and temperature on crenation of ghosts by

divalent and monovalent cationic salts. This appears plausible since it can explain the following observations.

1) Ghosts in 5 mM NaP<sub>i</sub> buffer, pH 8.0, 0°C were crenated by hemolytic isotonic salt 150 mM NH<sub>4</sub>HCO<sub>3</sub> or 1 mM 2,4-dinitrophenol (DNP), but ghosts isolated in this hemolytic salt at 0°C were globoid and not crenated by DNP [5]. It was previously shown that the band  $3_0$ /band  $3_1$ equilibrium ratio in erythrocytes was 0.1 in the selfexchange of Cl<sup>-</sup> at pH 7.8, 0°C, but this ratio was 10 in the self-exchange of  $HCO_3^{-1}$  [31]. The large difference between these two ratios does not appear to present an advantage in HCO<sub>3</sub><sup>-</sup> transport in blood, but with respect to the mechanism of control of the erythrocyte shape this difference would indicate that the Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> self-exchanges by band 3 are intrinsically echinocytogenic and stomatocytogenic, respectively. This is plausible since it provides an explanation for the Cl-dependent crenation of ghosts by 1 mM DNP and by 0.11 mM lysolecithin [5] as well as the appearance of ghosts as globoid and of an absence of crenation by 1 mM DNP isolated in hemolytic isotonic 150 mM NH<sub>4</sub>HCO<sub>3</sub>. The band  $3_0$ /band  $3_1$  equilibrium ratio in the self-exchange of divalent  $P_i$  with a H<sup>+</sup> by band 3 at pH 8.0, 0°C would probably to be similar to that of Cl<sup>-</sup>, since ghosts became crenated when the rate of this self-exchange was increased by decreasing the pH [5]; hence crenation of ghosts in 5 mM NaPi, pH 8.0, 0°C by isotonic 150 mM NH<sub>4</sub>HCO<sub>3</sub> and by 1 mM DNP. Crenation of ghosts in this buffer by isotonic 150 mM NH<sub>4</sub>HCO<sub>3</sub> would not be precluded by the opposition of  $HCO_3^-$  to crenation if the crenated ghosts were stabilized by dehydration by this salt [4]. Isolation of ghosts in this hemolytic salt at 0°C would not be accompanied by dehydration since the diffusion of this salt across the membrane would be accompanied by an increase of cell water. Ghosts isolated in hemolytic isotonic 300 mM glycerol at 0°C also had a globoid shape and were not crenated by isotonic 150 mM NaCl [5]. However, these ghosts are not equivalent to those isolated in hemolytic isotonic 150 mM NH<sub>4</sub>HCO<sub>3</sub> because of the destabilization of the skeleton in the absence of Cl<sup>-</sup> (see later), and ghosts isolated in 10 mM NaHCO<sub>3</sub>, pH 7.4, 0°C were as stable as those isolated in other 10 mM anionic buffers, pH 7.4, 0°C [22].

2) Crenation of ghosts in 10 mM Hepes buffer, pH 7.0, 4 °C occurred significantly only in presence of both 0.2 mM MgCl<sub>2</sub> and 10 mM NaCl [6]. Moreover, ghosts in 5 mM NaP<sub>i</sub> buffer, pH 8.0, 0°C were strongly crenated by 1 mM MgSO<sub>4</sub>, but those in 25-50 mM MgSO<sub>4</sub> were uncrenated [6]. These observations can be rationalized as follows. The selfexchanges of Cl<sup>-</sup> or divalent  $P_i$  with a H<sup>+</sup> by band 3 are intrinsically echinocytogenic. The divalent  $SO_4^{2-}$  can not be self-exchanged by band 3 at basic pH, in contrast to divalent  $P_i$ . However, divalent  $SO_4^{2^-}$  with a  $H^+$  can be self-exchanged by band 3 at acid pH, which would favor crenation of ghosts by  $Mg^{2+}$  [32]. An implication of a Cl or P<sub>i</sub> -dependent crenation by  $Mg^{2+}$  would be that this cation binds specifically at a site on spectrin, the major protein of the skeleton (see Fig. 2). The basis of the crenation of ghosts by other divalent cations appears to be the same, since crenation of ghosts in 10 mM Hepes buffer, pH 7.0, 4°C also became significant only in the presence of both chloride salts of these other divalent cations and NaCl at relatively low concentrations [6]. It is plausible that the basis of the crenation of ghosts by monovalent cationic salts at the relatively low concentration of 30 mM would be the same [4].

3) The dependence of Cl<sup>-</sup> on the skeleton conformation can also explain the spontaneous transformation into smooth discs and cups when the temperature was raised to 37 or 30°C after crenation by an isotonic saline of ghosts in 10 mM zwitterionic and cationic buffers, pH 7.0 or 7.4, 0-4°C [6, 22], since Cl<sup>-</sup> was absent or present at relatively low concentrations in these hypotonic buffers. This explanation is supported by the following observations. a) The irreversibility of the spontaneous transformation to discs and cups at 30°C of ghosts isolated in hypotonic zwitterionic buffers, pH 7.4, 0°C [22]. b) The stabilization at 37°C of ghosts isolated in hypotonic zwitterionic and cationic buffers, pH 7.4, 0°C supplemented with NaCl [9]. c) The previous unexpected observation that the skeleton in ghosts was less stable than that isolated at a very low ionic strength of the medium (very low concentrations of Cl<sup>-</sup> or P<sub>i</sub> in the medium) [33].

buffers containing Ghosts in relatively high concentrations of Cl<sup>-</sup> or P<sub>i</sub> were also unstable as indicated by their loss of the ability to be crenated by different agents over time depending of the pH and temperature [5, 17]. The agents included 10<sup>-6</sup>-10<sup>-7</sup> M Ca<sup>2+</sup>, 150 mM NaCl and 1 M NaCl plus 1 mM DNP. This instability would be attributed to destabilization of the skeleton as a result of band 3 losing the ability of transporting anions, as indicated by this observation. The rate of the loss of crenatability of ghosts in 5 mM NaP<sub>i</sub> buffer by 150 mM NaCl increased significantly as the pH increased from 6 to 8, as would be expected since band 3 transports divalent  $P_i$  with a  $H^+$  [5].

4) The minimal spontaneous transformation into smooth discs and cups observed when ghosts were isolated in zwitterionic and cationic buffers between pH 6.5-7.0, 0°C [22] could be due to the presence of sufficient amounts Cl<sup>-</sup> in these cationic buffers binding to band 3, or to the transport by band 3 of the anionic zwitterionic species with a  $H^+$  of zwitterionic buffers stabilizing the skeleton, as indicated by the following observations: a) The pKa of the 2 cationic buffers used, 2-[bis (-hydroxyethyl) amino]-2-(hydroxymethyl) propane-1,3-diol (Bistris) and imidazole, are 6.66 and 7.38 at 0°C, respectively [22]. b) Different zwitterionic buffers crenated erythrocytes at a low hematocrit (0.06 %) in a low ionic strength isotonic sucrose, pH 6.0, 20-22°C [34], which is likely due to their inward transport with a  $H^+$  by band 3, since the zwitterionic amino acid glycine and the bulky divalent band 3 high affinity inhibitor 4,4'-dinitro-2,2'-stilbenedisulfonate (DNDS) were transported with a  $H^+$ by band 3 at acid pH [35, 36]. Erythrocytes after crenation underwent a discocytosis followed by a cupping which may be due to the opposite transport of the buffer. c) Finally, the percentage of smooth discs and cups as a function of the pH of different zwitterionic buffers increased significantly when the pH increased from pH 7.0 to a certain pH, and then decreased significantly as the pH was further increased [33]. The transition of each of the zwitterionic buffers occurred at a pH which was consistent with its pK<sub>a</sub>, thus indicating that the skeleton was stabilized by the transport of its anionic species by band 3. However, the percentage of smooth discs and cups at pH 7.4 in a given zwitterionic buffer may be



Fig. (2). The basis of the crenation of ghosts by  $Mg^{2+}$ . Ghost crenation by  $Mg^{2+}$  occurs only in the self-exchange of Cl<sup>-</sup> by band 3, suggesting that  $Mg^{2+}$  binds at a specific site on spectrin. Other observations on ghosts cited in the text can be explained by self-exchanges of Cl<sup>-</sup>, HCO<sub>3</sub><sup>-</sup> or P<sub>1</sub> by band 3, a destabilization of the skeleton or an ability of band 3 of transporting zwitterionic buffers.

significantly different from that of another with the same  $pK_a$  (e.g. Tes (43 %) vs Hepes (100 %) (Fig. 2 of ref. 22). This difference may be due to binding to the transport site of band 3 of zwitterionic buffers with a lower percentage of smooth discs and cups with a lower steric hindrance.

In conclusion, the marked influence of ionic conditions and temperature on the crenation of ghosts by electrolytes can be explained by a mechanism of control of the erythrocyte shape, which would have the following major implications. a) The shape of ghosts isolated in hemolytic hypotonic buffers is specifically influenced by the type of the buffer and the pH. b) The instability of ghosts isolated in hypotonic zwitterionic and cationic buffers at some pH and temperature is the result of an absence of or of a relatively low concentration of Cl, which would destabilize the skeleton. c) The instability of ghosts in buffers containing a relatively high concentration of Cl<sup>-</sup> or P<sub>i</sub> is caused by a destabilization of the skeleton as a result of band 3 of losing the ability to transport anions. d) Crenation by divalent cations  $Mg^{2+}$  and  $Ca^{2+}$  depends on  $Cl^{-}$  thus suggesting that these cations crenate by binding at specific sites on spectrin, the major protein of the skeleton. The Cl<sup>-</sup> dependent crenation by these two divalent cations as well as by amphiphiles can be explained by the Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> selfexchanges by band 3 being intrinsically echinocytogenic and stomatocytogenic, respectively. e) Lastly, the ability of resealing ghosts under certain conditions in the presence of  $Mg^{2+}$  [1, 10, 11] is likely to be the result of a stabilization of the skeleton by a high Cl<sup>-</sup> concentration, a slow transport by band 3 of an anionic species of a buffer or of an inorganic salt and a specific binding of  $Mg^{2+}$  to a spectrin site.

#### **CONFLICT OF INTEREST**

The authors confirm that this article content has no conflicts of interest.

#### ACKNOWLEDGEMENTS

We thank Dr. MJ Whalley for reading the manuscript.

#### REFERENCES

- Schwoch G, Passow H. Preparation and properties of human erythrocyte ghosts. Mol Cell Biochem 1973; 2: 197-218.
- [2] Vaughan L, Penniston JT. Cation control of erythrocyte membrane shape: Ca<sup>++</sup> reversal of discocyte to echinocyte transition caused by Mg<sup>++</sup> and other cations. Biochem Biophys Res Commun 1976; 73: 200-5.
- [3] Johnson RM, Robinson J. Morphological changes in asymmetric erythrocyte membranes induced by electrolytes. Biochim Biophys Res Commun 1976; 70: 925-31.
- Johnson RM, Taylor G, Meyer DB. Shape and volume changes in erythrocyte ghosts and spectrin-actin networks. J Cell Biol 1980; 86: 371-6.
- [5] Lange Y, Gough A, Steck TL. Role of the bilayer in the shape of the isolated erythrocyte membrane. J Membr Biol 1982; 69: 113-23.
- [6] Sheetz MP. In: Revel JP, Henning U, Fox CF, Eds. Cell Shape and Surface Architecture, New York: Alan R. Liss 1977; pp. 559-67.
- [7] Patel VP, Fairbanks G. Spectrin phosphorylaton and shape change of human erythrocyte ghosts. J Cell Biol 1981; 88: 430-40.
- [8] Jinbu Y, Nakao M, Otsuka M et al. Two steps in ATP-dependent shape change of human erythrocyte ghosts. Biochem Biophys Res Commun 1983; 112: 384-90.
- [9] Pinteric L, Manery JF, Chaudry IH *et al.* The effect of EDTA, cations and various buffers on the morphology of erythrocyte membranes: an electron-microscopic study. Blood 1975; 45: 709-24.

- [10] Hoffman JF. Cation transport and structure of the red-cell plasma membrane. Circulation 1962; 26: 1202-13.
- [11] Steck TL, Kant JA. Preparation of impermeable ghosts and insideout vesicles from human erythrocyte membranes. Methods Enzymol 1974; 31 (pt A): 172-80.
- [12] Messana I, Orlando M, Cassiano L et al. Human erythrocyte metabolism is modulated by the O<sub>2</sub>-linked transition of hemoglobin. FEBS Lett 1996; 390: 25-8.
- [13] Campanella ME, Chu H, Low PS. Assembly and regulation of a glycolytic enzyme complex on the human erythrocyte membrane. Proc Nat Acad Sci USA 2005; 102: 2402-7.
- [14] Tuvia S, Levin S, Korenstein R. Oxygenation-deoxygenation cycle of erythrocytes modulates submicron cell membranes fluctuations. Biophys J 1992; 63: 599-602.
- [15] Palek J, Stewart G, Lionetti FJ. The dependence of shape of human erythrocyte ghosts on calcium, magnesium, and adenosine triphosphate. Blood 1974; 44: 583-97.
- [16] Jinbu Y, Sato S, Nakao T *et al.* The role of ankyrin in shape and deformability change of human erythrocyte ghosts. Biochem Biophys Acta 1984; 773: 237-45.
- [17] Jinbu Y, Sato S, Nakao M et al. Ca<sup>2+</sup> and Mg-ATP-dependent shape change of human erythrocyte ghosts and triton shells. Exp Cell Res 1984; 151: 160-70.
- [18] Seigneuret M, Devaux PF. ATP-dependent asymmetric distribution of spin-labeled phospholipids in the erythrocyte membrane: relation to shape changes. Proc Natl Acad Sci USA 1984; 81: 3751-
- [19] Herrmann A, Müller P, Glaser R. Shape transformations of erythrocyte ghosts depends on ion concentrations. Biosci Rep 1985; 5: 417-23.
- [20] Patel VP, Fairbanks G. Relationship of major phosphorylation reactions and MgATPase activities to ATP-dependent shape change of human erythrocyte membranes. J Biol Chem 1986; 261: 3170-7.
- [21] Schrier SL, Junga I, Ma L. Studies on the effect of vanadate on endocytosis and shape changes in human red blood cells and ghosts. Blood 1986; 68: 1008-14.
- [22] Raval PJ, Carter DP, Fairbanks G. Relationship of hemolysis buffer structure, pH and ionic strength to spontaneous contour smoothing of isolated erythrocyte membranes. Biochim Biophys Acta 1989; 983: 230-40.

Revised: October 07, 2013

Accepted: October 08, 2013

© Pierre Wong et al.; Licensee Bentham Open.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/), which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.

- [23] DeLoach JR, Droleskey R. Preparation of ovine carrier erythrocytes: their action and survival. Comp Biochem Physiol 1986; 84A: 441-5.
- [24] Sanz S, Lizano C, Luque J et al. In vitro and in vivo study of glutamate dehydrogenase encapsulated into mouse erythrocytes by a hypotonic dialysis procedure. Life Sci 1999; 65: 2781-9.
- [25] Milanick MA, Ritter S, Meissner K. Engineering erythrocytes to be electrosensors: first steps. Blood Cells Mol Dis 2011; 47: 100-6.
- [26] Wong P. A basis of echinocytosis and stomatocytosis in the discsphere transformations of the erythrocyte. J Theor Biol 1999; 196: 343-61.
- [27] Cabantchik ZI, Greber R. Chemical probes for anion transporters of mammalian cell membranes. Am J Physiol (Cell Physiol) 1992; 262: C803-27.
- [28] Wong P. The basis of echinocytosis of the erythrocyte by glucose depletion. Cell Bioch Funct 2011; 29: 708-11.
- [29] Hayashi Y, Katsumoto Y, Oshige I et al. Dielectric inspection of erythrocytes. J Non-Cryst Sol 2010; 356: 757-62.
- [30] Weast RC, Astle MJ, Eds., CRC Hanbook of Chemistry and Physics 59<sup>th</sup> ed, CRC Press Inc. West Palm Beach, Florida, 1978-1979, p. D-204.
- [31] Knauf PA, Law FY, Leung TWV, et al. Substrate-dependent reversal of anion transport site orientation in the human red blood cell anion-exchange protein, AE1. Proc Natl Acad Sci USA 2002; 99: 10861-4.
- [32] Milanick MA, Gunn RB. Proton-sulfate cotransport: external protonation activation of sulphate influx into human red blood cells. Am J Physiol (Cell Physiol) 1984; 247: C247-59.
- [33] Lange Y, Hadesman RA, Steck TL. Role of the reticulum in the stability and shape of the isolated human erythrocyte membrane. J Cell Biol 1982; 92: 714-21.
- [34] Rudenko SV. Characterization of morphological response of red cells in a sucrose solution. Blood Cells Mol Dis 2009; 42: 252-61.
- [35] King PA, Gunn RB. Glycine transport by human red blood cells and ghosts: evidence for glycine anion and proton cotransport by band 3. Am J Physiol (Cell Physiol) 1991; 261: C814-21.
- [36] Salhany JM, Cordes KS, Sloan RL, Band 3 (AE1, SLCA1)mediated transport of stilbenedisulfonates. I: Functional identification of the proton-activated stilbenedisulfonate influx site. Blood Cells Mol Dis 2006; 37: 137-48.

Received: September 13, 2013